

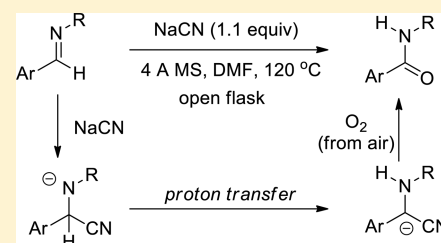
Formation of Amides from Imines via Cyanide-Mediated Metal-Free Aerobic Oxidation

Hong-Ahn Seo,[‡] Yeon-Ho Cho,[‡] Ye-Sol Lee, and Cheol-Hong Cheon*[‡]

Department of Chemistry, Korea University, 145 Anam-ro, Seongbuk-gu, Seoul 02841, Republic of Korea

S Supporting Information

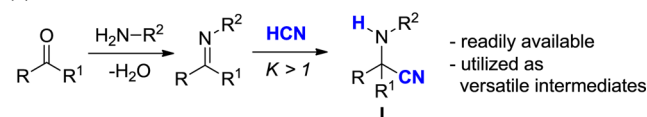
ABSTRACT: A new protocol for the direct formation of amides from imines derived from aromatic aldehydes via metal-free aerobic oxidation in the presence of cyanide is described. This protocol was applicable to various aldimines, and the desired amides were obtained in moderate to good yields. Mechanistic studies suggested that this aerobic oxidative amidation might proceed via the addition of cyanide to imines followed by proton transfer from carbon to nitrogen in the original imines, leading to carbanions of α -amino nitriles, which undergo subsequent oxidation with molecular oxygen in air to provide the desired amide compounds.



INTRODUCTION

Ever since Strecker's original report in 1850 on the three-component reaction between aldehydes, ammonia, and hydrogen cyanide (HCN),¹ the addition of HCN to imines derived from carbonyls and amines leading to α -aminonitriles has been the subject of great interest because the resulting Strecker products **I** have been widely used as versatile intermediates in a number of synthetic applications (Figure 1a).^{2,3} Although a

(a) addition of HCN to imines



(b) addition of CN⁻ to imines

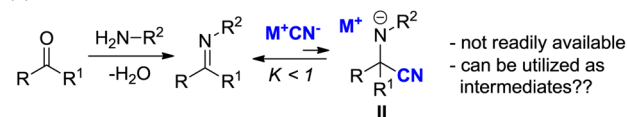


Figure 1. Comparison of the addition products of HCN and CN⁻ to imines.

similar addition of cyanide anion (CN⁻) to imines has been known for a long time, synthetic protocols utilizing the resulting cyanide adducts **II** as synthetic intermediates have been far less developed than those with HCN adducts **I**. This might be due to the fact that cyanide adducts **II** readily undergo an elimination reaction (retro-Strecker reaction), which makes it difficult to maintain high concentration of adducts **II** so that the rate of their conversion to the next step is reasonably high (Figure 1b).⁴

Recently, our group initiated a program to develop novel synthetic protocols utilizing cyanide adducts **II** as valuable synthetic intermediates. For example, we developed a protocol for the synthesis of benzoxazoles from *o*-aminophenol and aldehydes via metal-free aerobic oxidative cyclization using

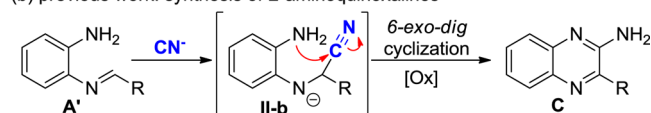
cyanide as the catalyst (Scheme 1a).⁵ Imine **A** is unlikely to undergo cyclization because 5-endo-trig cyclization is disfa-

Scheme 1. Synthetic Applications of Cyanide Adducts **II** to Organic Transformations via Aerobic Oxidation

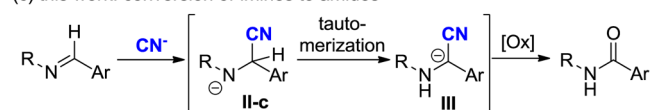
(a) previous work: synthesis of benzoxazoles



(b) previous work: synthesis of 2-aminoquinoxalines



(c) this work: conversion of imines to amides



vored according to Baldwin's rules.⁶ However, cyanide adduct **II-a** readily underwent cyclization through favored 5-*exo*-tet cyclization and subsequent aerobic oxidation of the cyclized product provided benzoxazole **B**. When we attempted to extend this method to the synthesis of benzimidazoles with *o*-phenylenediamine, intermediate **II-b** underwent 6-*exo*-dig cyclization rather than the expected 5-*exo*-tet cyclization, leading to the formation of 2-aminoquinoxaline **C** after aerobic oxidation (Scheme 1b).⁷

Herein we report the development of another synthetic protocol using cyanide adducts **II** as the intermediates: the

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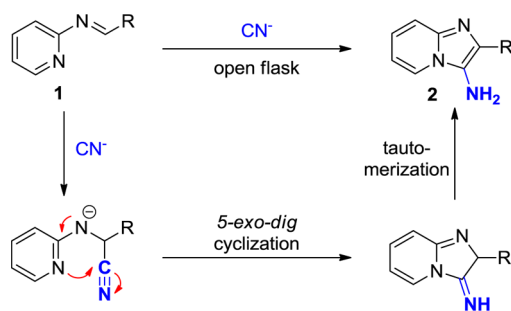
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direct formation of amides from imines via metal-free aerobic oxidation (Scheme 1c).^{8–11} This result is based on our recent finding of the unexpected formation of amides from aromatic aldimines in the presence of cyanide, when we attempted to utilize cyanide as a surrogate for isocyanide in the synthesis of imidazopyridines. Mechanistic studies suggested that cyanide adducts II-c underwent tautomerization to generate benzylic anions III,¹² and subsequent aerobic oxidation of III provided the corresponding amides.

RESULTS AND DISCUSSION

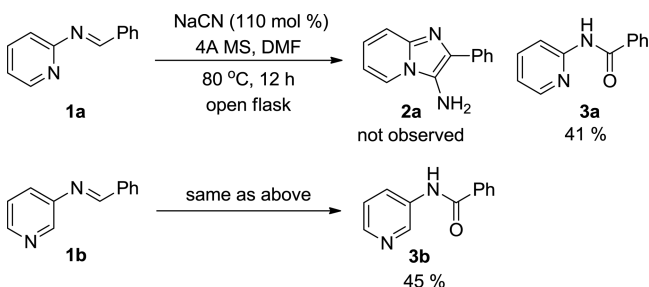
Because we demonstrated that cyanide could be utilized as a surrogate for isocyanide in the synthesis of 2-aminoquinoxalines (Scheme 1b),^{7,13} we attempted to extend the usage of cyanide as a surrogate for isocyanide to the synthesis of other nitrogen-containing heteroaromatic compounds, such as imidazopyridine 2.¹⁴ For example, it was expected that imidazopyridine 2 could be prepared from imine 1, derived from 2-aminopyridine and aldehyde, using cyanide as a surrogate for isocyanide via addition of cyanide to imine 1 followed by 5-exo-dig cyclization (Scheme 2).

Scheme 2. Application of CN⁻ as a Surrogate for Isocyanide in the Synthesis of Imidazopyridine 2



On the basis of this idea, imine 1a, derived from 2-aminopyridine and benzaldehyde, was subjected to the aerobic oxidative cyclization conditions⁷ previously used for the synthesis of 2-aminoquinoxalines. However, the expected imidazopyridine 2a was not observed; rather unexpectedly, amide 3a was obtained as the major product (Scheme 3).¹⁰

Scheme 3. Unexpected Formation of Amides from Imines



Because the direct conversion of imines to amides via aerobic oxidation in the presence of cyanide has been rarely reported in the literature even with the assistance of metal catalysts,^{11,15} we initially suspected that the nitrogen atom in the pyridine ring of 1a might play a role in this aerobic oxidative amidation reaction. Thus, we tested the same transformation with imine 1b, which is derived from 3-aminopyridine instead of 2-

aminopyridine. When 1b was subjected to the above conditions, the formation of the corresponding amide 3b was again observed.

Because these results strongly suggested that the nitrogen atom in the pyridine ring in imine 1 is not required for the formation of amides from imines, the reaction conditions were optimized using imine 4a, derived from aniline and benzaldehyde, as a model compound (Table 1). Cyanide was

Table 1. Optimization of Reaction Conditions

entry	NaCN (x mol %)	solvent	temp (°C)	time (h)	yield of 5a (%) ^a
1	110	DMF	80	4	69
2	–	DMF	80	24	N.R. ^b
3 ^c	110	DMF	80	24	<5
4	110	DMSO	80	4	56
5	110	CH ₃ CN	80	24	40
6	110	dioxane	80	24	N.R. ^b
7	110	toluene	80	24	N.R. ^b
8	110	<i>i</i> -PrOH	80	24	N.R. ^b
9	50	DMF	80	8	35
10	200	DMF	80	4	74
11	300	DMF	80	4	70
12	110	DMF	100	2	71 (13) ^d
13	110	DMF	120	1	70 (12) ^d
14	110	DMF	120	2	81 ^e
15 ^f	110	DMF	120	2	79

^aIsolated yield. ^bNo reaction. ^cUnder an argon atmosphere. ^dThe number in parentheses is the isolated yield of 6a. ^eThe isolated yield of 5a after basic hydrolysis. ^fSequential one-pot protocol.

found to play a crucial role in this protocol; amide 5a was obtained in good yield in the presence of cyanide, whereas the amide formation did not occur at all in the absence of cyanide (entries 1 and 2). When this reaction was performed under an argon atmosphere, a negligible amount of 5a was observed, indicating that molecular oxygen in air is the terminal oxidant in this transformation (entry 3). The choice of solvent was found to have a strong influence on the formation of 5a (entries 1 and 4–8). Among the solvents tested, 5a was obtained in the best yield in DMF, and thus DMF was chosen for further investigations.

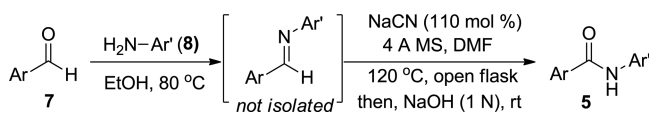
Next, the effect of the amount of cyanide on the formation of 5a was examined (entries 1, 2, and 9–11). The amount of cyanide was found to exert a strong influence on the efficacy of this transformation. The yield of 5a increased with the amount of cyanide until a stoichiometric amount of cyanide was used. However, the yield of 5a did not improve further and reached a plateau when superstoichiometric amounts of cyanide were used. Thus, we decided to use a slight excess of cyanide (110 mol %) as the optimal amount.

Despite numerous effects, we were not able to further improve the yield of 5a higher than 70% yield, and a careful analysis of the reaction mixture revealed that α -imino nitrile 6a¹⁶ was also formed albeit in low yield. Because α -imino nitriles could be converted to the corresponding amides via hydrolysis,¹⁷ we attempted to convert 6a into 5a at an elevated reaction temperature (entries 1, 12, and 13). Rather

disappointingly, the yield of **5a** did not improve, and **6a** was still obtained in similar yields even at higher temperatures, although the reaction rate accelerated at high temperature. However, when the reaction mixture was treated with a NaOH solution after complete consumption of **4a**, **5a** was obtained in 81% yield (entry 14). Finally, we further developed a one-pot protocol for the synthesis of **5a** directly from the aldehyde and aniline without isolation of imine **4a**. When **4a**, prepared in situ from benzaldehyde and aniline, was subjected to the standard aerobic oxidation conditions, the yield of **5a** was similar to that obtained using isolated imine **4a**, after treatment of the reaction mixture with an aqueous basic solution (entry 15).¹⁸

With these optimized conditions in hand, we explored the substrate scope for this transformation (Table 2). Various

Table 2. Substrate Scope



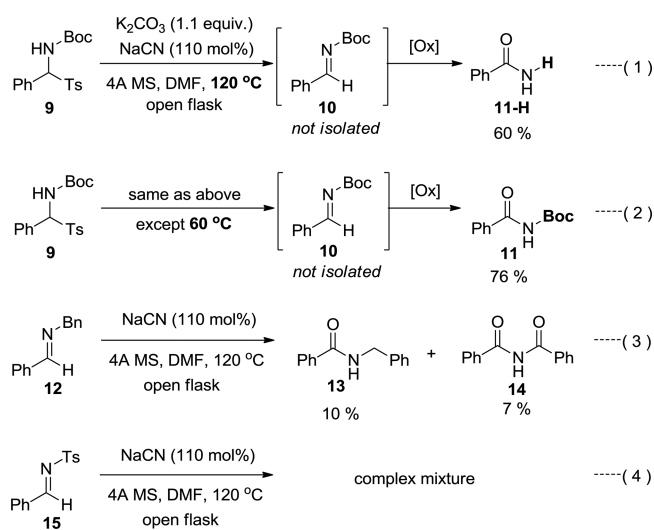
entry	amide (5)	Ar	Ar'	yield (%) ^a
1	5a	C ₆ H ₅	Ph	79
2	5b	4-MeOC ₆ H ₄	Ph	76
3	5c	4-MeC ₆ H ₄	Ph	67
4	5d	4-ClC ₆ H ₄	Ph	60
5	5e	4-BrC ₆ H ₄	Ph	57
6	5f	4-MeCO ₂ C ₆ H ₄	Ph	77
7	5g	2-MeOC ₆ H ₄	Ph	71
8	5h	2-MeC ₆ H ₄	Ph	81
9	5i	2-ClC ₆ H ₄	Ph	64
10	5j	1-naphthyl	Ph	69
11	5k	2-naphthyl	Ph	56
12	5l	2-furyl	Ph	48
13	5m	2-thienyl	Ph	71
14	5n	2-pyridinyl	Ph	83
15	5o	C ₆ H ₅	4-BrC ₆ H ₄	65
16	5p	C ₆ H ₅	4-MeOC ₆ H ₄	62

^aIsolated yield.

aromatic aldehydes could be applied to this protocol, and amides **5** were obtained in good yields (entries 1–9). The electronic properties of the aldehydes exerted some influence on this transformation; aldehydes bearing electron-donating groups (entries 2 and 3) provided the corresponding amides in better yields than those with electron-withdrawing substituents (entries 4–6). More sterically hindered ortho-substituted aldehydes were also amenable to this protocol, and the corresponding amides **5** were obtained in good yields (entries 7–9). This protocol was further extended to fused aromatic aldehydes and heteroaromatic aldehydes, and the desired products were obtained in moderate to good yields (entries 10–14). Next, we investigated the effect of the electronic properties of aniline derivatives on this transformation (entries 1, 15 and 16). The electronic nature of anilines was found to have only a small influence on the yield of this transformation; aniline derivatives bearing either electron-rich or electron-deficient substituents provided the desired products in slightly lower yields than aniline itself.

We further investigated other imines derived from amines different from anilines for this aerobic oxidative amidation reaction (Scheme 4). When a precursor **9** of Boc imine **10** was subjected to the standard conditions in the presence of K₂CO₃,

Scheme 4. Investigation of Amine Sources

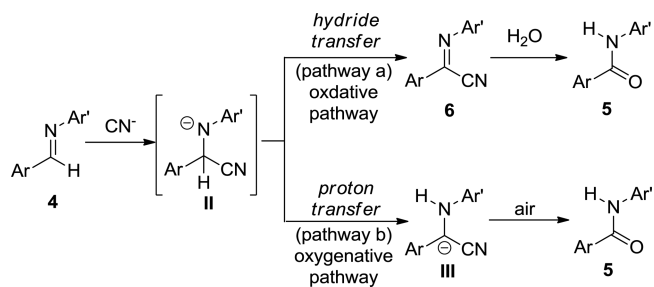


the corresponding in situ-generated Boc imine **10** readily underwent oxidative amidation, but the Boc group in the resulting amide **11** was removed to provide benzamide **11-H** in 60% yield (eq 1).

However, when the same transformation was performed at 60 °C, the desired Boc protected amide **11** was obtained in 76% yield without any concomitant formation of **11-H** (eq 2).²⁰ Unfortunately, however, other imines derived from other amine sources, such as benzyl amine and tosyl amine, were found not to be applicable to this aerobic oxidative amidation; benzyl imine **12** afforded the corresponding amide **13** and imide **14** only in low yields (~10%) (eq 3), while tosyl imine **15** provided a complex mixture (eq 4).

With these results in hand, we attempted to elucidate the reaction mechanism for this cyanide-mediated aerobic oxidative amidation of imines. As cyanide was found to play a critical role in this transformation, cyanide adduct **II** might be the key intermediate. From intermediate **II**, there might be two possible reaction pathways for the aerobic oxidative amidation reaction (Scheme 5).¹⁹

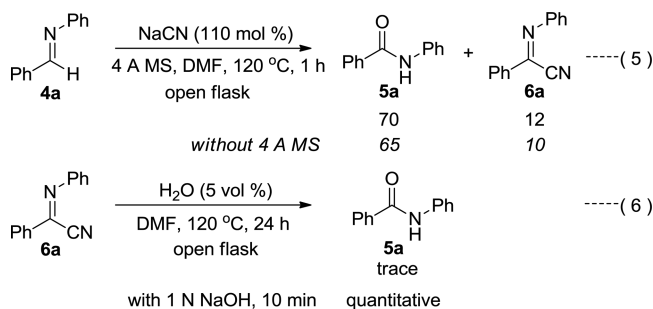
Scheme 5. Possible Reaction Pathways



Intermediate **II** might first undergo oxidation through hydride transfer to form α -imino nitrile **6**, with subsequent hydrolysis of **6** affording the desired amide **5** (pathway a; oxidative pathway). On the other hand, proton transfer from the α -carbon atom to the nitrogen atom in intermediate **II** might take place to generate anion **III**,¹² which subsequently undergoes reaction with molecular oxygen to furnish amide **5** (pathway b; oxygenative pathway).^{18,19b}

Because the formation of α -imino nitriles **6** was observed during the optimization of the reaction conditions, we initially suspected that amides **5** might be formed through hydrolysis of α -imino nitriles **6**.¹⁷ Under such a scenario, it was expected that water would play a role in this transformation. To test this idea, we carried out a few controlled experiments (Scheme 6). When

Scheme 6. Test of Iminonitrile 4a as an Intermediate in the Aerobic Oxidative Amidation



the reaction was performed in the absence of molecular sieves (i.e., in the presence of water), the yield of **5a** was not improved, and **5a** and **6a** were obtained in similar level of yields regardless of the presence of molecular sieves (eq 5). Furthermore, when the isolated α -imino nitrile **6a** was directly subjected to hydrolysis in wet organic solvent, **6a** did not undergo the hydrolysis at 120 °C even after 24 h (eq 6). Instead, strong basic conditions (1 N NaOH) were needed to hydrolyze α -imino nitrile **6a** into amide **5a**. These results led us to rule out the possibility that the amides would be formed via the hydrolysis of α -imino nitriles even though **6** was observed in the reaction mixture. On the basis of these experimental results in conjunction with the previous reports on dimerization of aldimines where intermediate **III** was considered to be formed from cyanide adduct **II** via tautomerization,^{12,15} we believe that the formation of amides from imines in the presence of cyanide would proceed via pathway b.

CONCLUSIONS

We report the direct formation of amides from imines in the presence of cyanide via metal-free aerobic oxidation. Various aromatic aldehydes were applicable to this protocol, and the desired amides were obtained in good to high yields. Furthermore, other imines derived from amine sources other than aniline could be applied to this protocol although the yields of the corresponding amides showed a dependence on the choice of amines. Mechanistic studies suggested that this aerobic oxidative amidation might proceed via the addition of cyanide to imines, followed by the proton transfer leading to carbanions of α -aminonitriles, and subsequent aerobic oxidation. The elucidation of the detailed reaction mechanism and further application of this method are currently underway in our laboratory.

EXPERIMENT SECTION

General. All reactions were carried out in oven- or flame-dried glassware in an open flask unless otherwise noted. Except as otherwise indicated, all reactions were magnetically stirred and monitored by analytical thin layer chromatography (TLC) using precoated silica gel glass plates (0.25 mm) with F254 indicator. Visualization was accomplished by UV light (254 nm), with combination of potassium permanganate and/or phosphomolybdic acid solution as an indicator.

Flash column chromatography was performed using silica gel 60 (230–400 mesh). Yields refer to chromatographically and spectroscopically pure compounds, unless otherwise noted. Commercial grade reagents and solvents were used without further purification. Liquid aldehydes were purified by fractional distillation, and solid aldehydes were purified by a column chromatography on silica. ¹H NMR spectra were recorded on either a 300 or a 400 MHz spectrometer. Tetramethylsilane (δ : 0.0 ppm) was used as an internal standard for ¹H NMR. The proton spectra are reported as follows: δ (position of proton, multiplicity, coupling constant *J*, number of protons). Multiplicities are indicated by s (singlet), d (doublet), t (triplet), q (quartet), p (quintet), h (septet), m (multiplet), and br (broad).

General Procedure for Metal-Free Aerobic Oxidative Amidation (Table 2). To a 50 mL two-necked round-bottomed flask equipped with a condenser were added amine **8** (1.1 mmol, 1.1 equiv) and ethanol (10 mL). To the above solution was added aryl aldehyde **7** (1.0 mmol, 1.0 equiv), and the reaction mixture was stirred at 80 °C and monitored by TLC. After complete consumption of aldehyde **7**, the reaction mixture was concentrated under reduced pressure to afford the crude product of imine. Without further purification of the resulting imine, the crude mixture was dissolved in DMF (10 mL), and sodium cyanide (1.1 mmol, 1.1 equiv) and molecular sieves were added to the reaction mixture. Then the reaction mixture was stirred at 120 °C until all the imine was completely consumed. Upon complete consumption of the imine, NaOH solution (1.0 N) was added to the reaction mixture and the reaction mixture was stirred at room temperature for 10 min. The crude mixture was extracted with ether, and the organic layer was combined, dried over MgSO₄, and concentrated. The resulting crude mixture was purified by flash column chromatography on silica to provide the expected amide **5**.

N-(Pyridin-2-yl)benzamide (3a). The desired product **3a** was obtained as a yellow solid (81 mg, 41% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:2): *R_f* = 0.50 (ethyl acetate/hexanes = 2:1). The spectroscopic data were in good agreement with the literature.²¹ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.77 (br, 1H), 8.38 (d, *J* = 4.1 Hz, 1H), 8.18 (d, *J* = 8.3 Hz, 1H), 8.02 (d, *J* = 8.0 Hz, 2H), 7.83 (t, *J* = 7.7 Hz, 1H), 7.46–7.62 (m, 4H).

N-(Pyridin-3-yl)benzamide (3b). The desired product **3b** was obtained as a yellow solid (85 mg, 45% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:4): *R_f* = 0.30 (ethyl acetate/hexanes = 1:4). The spectroscopic data were in good agreement with the literature.²² ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.45 (br, 1H), 8.92 (d, *J* = 1.9 Hz, 1H), 8.30 (d, *J* = 4.7 Hz, 1H), 8.20 (d, *J* = 8.0 Hz, 1H), 7.96 (d, *J* = 7.2 Hz, 2H), 7.52–7.63 (m, 3H), 7.36–7.41 (m, 1H).

N-Phenylbenzamide (5a). The desired product **5a** was obtained as a white solid (147 mg, 79% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R_f* = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.24 (br, 1H), 7.94 (d, *J* = 7.2 Hz, 2H), 7.77 (d, *J* = 8.3 Hz, 2H), 7.48–7.62 (m, 3H), 7.34 (t, *J* = 7.7 Hz, 2H), 7.09 (t, *J* = 7.2 Hz, 1H).

4-Methoxy-N-phenylbenzamide (5b). The desired product **5b** was obtained as a yellow solid (173 mg, 76% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R_f* = 0.40 (ethyl acetate/hexanes = 1:3). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.07 (br, 1H), 7.94 (d, *J* = 8.3 Hz, 2H), 7.75 (d, *J* = 7.7 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.01–7.10 (m, 3H), 3.82 (s, 3H).

4-Methyl-N-phenylbenzamide (5c). The desired product **5c** was obtained as a white solid (133 mg, 67% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R_f* = 0.40 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, CDCl₃, ppm): δ 7.77 (d, *J* = 8.0 Hz, 2H), 7.64 (d, *J* = 7.7 Hz, 2H), 7.37 (t, *J* = 7.7 Hz, 2H), 7.29 (t, *J* = 7.7 Hz, 2H), 7.15 (t, *J* = 7.3 Hz, 1H), 2.43 (s, 3H).

4-Chloro-N-phenylbenzamide (5d). The desired product **5d** was obtained as a white solid (139 mg, 60% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R_f* = 0.30

(ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, CDCl₃, ppm): δ 7.82 (d, *J* = 8.2 Hz, 2H), 7.73 (br, 1H), 7.62 (d, *J* = 7.8 Hz, 2H), 7.48 (d, *J* = 8.6 Hz, 2H), 7.39 (t, *J* = 7.6 Hz, 2H), 7.17 (t, *J* = 7.4 Hz, 1H).

4-Bromo-*N*-phenylbenzamide (5e). The desired product **5e** was obtained as a yellow solid (133 mg, 57% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.75 (d, *J* = 8.2 Hz, 2H), 7.63 (dd, *J* = 7.8 Hz, *J* = 4.7 Hz, 4H), 7.39 (t, *J* = 7.8 Hz, 2H), 7.15–7.20 (m, 1H).

4-Methoxycarbonyl-*N*-phenylbenzamide (5f). The desired product **5f** was obtained as a yellow solid (196 mg, 77% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:4): *R*_f = 0.30 (ethyl acetate/hexanes = 1:4). The spectroscopic data were in good agreement with the literature.²⁴ ¹H NMR (400 MHz, CDCl₃, ppm): δ 8.16 (d, *J* = 7.8 Hz, 2H), 7.94 (d, *J* = 7.8 Hz, 2H), 7.82 (br, 1H), 7.65 (d, *J* = 7.8 Hz, 2H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.19 (m, 1H), 3.97 (s, 3H).

2-Methoxy-*N*-phenylbenzamide (5g). The desired product **5g** was obtained as a white solid (161 mg, 71% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²⁴ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.10 (br, 1H), 7.72 (d, *J* = 8.0 Hz, 2H), 7.62 (d, *J* = 8.0 Hz, 1H), 7.49 (t, *J* = 7.7 Hz, 1H), 7.32 (t, *J* = 7.4 Hz, 2H), 7.17 (d, *J* = 8.3 Hz, 1H), 7.01–7.10 (m, 2H), 3.88 (s, 3H).

2-Methyl-*N*-phenylbenzamide (5h). The desired product **5h** was obtained as a white solid (171 mg, 81% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:8): *R*_f = 0.60 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.28 (br, 1H), 7.73 (d, *J* = 8.0 Hz, 2H), 7.24–7.47 (m, 6H), 7.07 (t, *J* = 7.3 Hz, 1H), 2.37 (s, 3H).

2-Chloro-*N*-phenylbenzamide (5i). The desired product **5i** was obtained as a yellow solid (148 mg, 64% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (400 MHz, CDCl₃, ppm): δ 7.89 (br, 1H), 7.76 (d, *J* = 7.4 Hz, 1H), 7.65 (d, *J* = 8.2 Hz, 2H), 7.35–7.48 (m, 5H), 7.18 (t, *J* = 7.2 Hz, 1H).

***N*-Phenyl-1-naphthalamide (5j).** The desired product **5j** was obtained as a white solid (170 mg, 69% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.57 (br, 1H), 8.13–8.20 (m, 1H), 7.99–8.10 (m, 2H), 7.71–7.83 (m, 3H), 7.55–7.63 (m, 3H), 7.36 (t, *J* = 7.8 Hz, 2H), 7.11 (t, *J* = 7.3 Hz, 1H).

***N*-Phenyl-2-naphthalamide (5k).** The desired product **5k** was obtained as a yellow solid (139 mg, 56% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²⁵ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.43 (br, 1H), 8.57 (s, 1H), 7.98–8.11 (m, 4H), 7.82 (d, *J* = 8.0 Hz, 2H), 7.59–7.66 (m, 2H), 7.37 (t, *J* = 7.3 Hz, 2H), 7.11 (t, *J* = 7.0 Hz, 1H).

***N*-Phenylfuran-2-carboxamide (5l).** The desired product **5l** was obtained as a dark brown solid (90 mg, 48% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:4): *R*_f = 0.30 (ethyl acetate/hexanes = 1:4). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.17 (br, 1H), 7.92 (s, 1H), 7.73 (d, *J* = 8.3 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 3H), 7.10 (t, *J* = 7.4 Hz, 1H), 6.67–6.71 (m, 1H).

***N*-Phenylthiophene-2-carboxamide (5m).** The desired product **5m** was obtained as a yellow solid (144 mg, 71% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.30 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²⁵ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.21 (br, 1H), 8.01 (d, *J* = 3.6 Hz, 1H), 7.84 (d, *J* = 5.0 Hz,

1H), 7.70 (d, *J* = 8.0 Hz, 2H), 7.34 (t, *J* = 7.7 Hz, 2H) 7.19–7.24 (m, 1H) 7.09 (t, *J* = 7.3 Hz, 1H).

***N*-Phenylpicolinamide (5n).** The desired product **5n** was obtained as a yellow solid (164 mg, 83% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:3): *R*_f = 0.40 (ethyl acetate/hexanes = 1:3). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.62 (br, 1H), 8.73 (d, *J* = 4.1 Hz, 1H), 8.12–8.20 (m, 1H), 8.06 (t, *J* = 7.6 Hz, 1H), 7.90 (d, *J* = 8.0 Hz, 2H), 7.62–7.70 (m, 1H), 7.35 (t, *J* = 7.7 Hz, 2H), 7.11 (t, *J* = 7.2 Hz, 1H).

***N*-(4-Bromophenyl)benzamide (5o).** The desired product **5o** was obtained as a yellow solid (176 mg, 65% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:8): *R*_f = 0.50 (ethyl acetate/hexanes = 1:5). The spectroscopic data were in good agreement with the literature.²⁶ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.36 (br, 1H), 7.93 (d, *J* = 7.7 Hz, 2H), 7.76 (d, *J* = 8.8 Hz, 2H), 7.49–7.62 (m, 5H).

***N*-(4-Methoxyphenyl)benzamide (5p).** The desired product **5p** was obtained as a yellow solid (141 mg, 62% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:4): *R*_f = 0.40 (ethyl acetate/hexanes = 1:3). The spectroscopic data were in good agreement with the literature.²³ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.12 (br, 1H), 7.93 (d, *J* = 6.9 Hz, 2H), 7.66 (d, *J* = 8.8 Hz, 2H), 7.47–7.59 (m, 3H), 6.91 (d, *J* = 9.1 Hz, 2H), 3.73 (s, 3H).

Benzamide (11-H). The desired product **11-H** was obtained as a white solid (72 mg, 60% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:1): *R*_f = 0.30 (ethyl acetate/hexanes = 1:1). The spectroscopic data were in good agreement with the literature.^{9b} ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 7.96 (br, 1H), 7.85 (d, *J* = 7.4 Hz, 1H), 7.47–7.53 (m, 1H), 7.40–7.47 (m, 2H), 7.35 (br, 1H).

***N*-(tert-Butyloxycarbonyl)benzamide (11).** The desired product **11** was obtained as a white solid (165 mg, 76% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:3): *R*_f = 0.30 (ethyl acetate/hexanes = 1:3). The spectroscopic data were in good agreement with the literature.²⁰ ¹H NMR (300 MHz, DMSO-*d*₆, ppm): δ 10.66 (br, 1H), 7.82 (d, *J* = 7.7 Hz, 2H), 7.54–7.61 (m, 1H), 7.42–7.50 (m, 2H), 1.46 (s, 9H).

***N*-Benzylbenzamide (13).** The desired product **13** was obtained as a white solid (21 mg, 10% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.70 (ethyl acetate/hexanes = 1:1). The spectroscopic data were in good agreement with the literature.^{9a} ¹H NMR (300 MHz, CDCl₃, ppm): δ 7.74–7.84 (m, 2H), 7.29–7.53 (m, 8H), 6.46 (br, 1H), 4.65 (d, *J* = 5.5 Hz, 2H).

***N*,2-Diphenylacetamide (14).** The desired product **14** was obtained as a white solid (15 mg, 7% yield) after column chromatography on silica (ethyl acetate/hexanes = 1:5): *R*_f = 0.60 (ethyl acetate/hexanes = 1:1). The spectroscopic data were in good agreement with the literature.²⁰ ¹H NMR (300 MHz, CDCl₃, ppm): δ 8.90 (br, 1H), 7.88 (d, *J* = 7.4 Hz, 4H), 7.58–7.67 (m, 2H), 7.52 (t, *J* = 7.6 Hz, 4H).

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b01922.

NMR spectra of all compounds (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*Phone: +82-2-3290-3147. Fax: +82-2-3290-3121. E-mail: cheon@korea.ac.kr.

Author Contributions

‡These authors contributed equally.

Notes

The authors declare no competing financial interests.

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